Monatshefte für Chemie Chemical Monthly Printed in Austria

Review

Thiohydantoin Nucleosides. Synthesis Approaches

Ahmed I. A. Khodair^{1,*,#}, El Sayed H. El Ashry², and Najim A. L. Al-Masoudi^{3,*,§}

- ¹ Chemistry Department, Faculty of Education, Tanta University (Kafr El-Sheikh Branch), Kafr El-Sheikh, Egypt
- ² Chemistry Department, Faculty of Science, University of Alexandria, Alexandria, Egypt
- ³ Chemistry Department, University of Konstanz, P.O. Box 5560, D-78457 Konstanz, Germany

Received January 8, 2004; accepted January 8, 2004 Published online June 24, 2004 © Springer-Verlag 2004

Summary. Several thiohydantoin *N*-nucleosides and their *S*-glycosides were prepared *via* different approaches which involved the direct glycosylation of the free thiohydantoin bases or their silylated derivatives with the corresponding sugar moieties in the presence of base or *Lewis* acid. Deprotection was carried out in acidic or basic medium. The site of *N*- and/or *S*-glycosylation was determined by NMR and UV measurements. In similar manner, hydantoin nucleosides were prepared.

Keywords. Biological activity; Glycosylation; Sugar derivatives; Pyrimidines; Hydantoin and thiohydantoin nucleosides.

Introduction

Hydantoin (1) was isolated first by *Bayer* [1] in the course of his classic research on uric acid (1861). The 2-thio analogue 2 was reported in 1890 [2]. The family of these compounds have been investigated intensively in connection with the structural activity of peptides, pencillin [3], 4-thiohydantoin (3) [4], 2,4-dithiohydantoin (4) [5], and their alkyl and aryl derivatives (Scheme 1). Reviews on the chemistry of hydantoins (2,4-imidazolidinediones) including thiohydantoins appeared successively in 1950 [6], 1957 [7], 1966 [8], and 1985 [9]. *Edward*'s review [8] dealt only with thiohydantoins, while *Ware*'s review [6] was particularly exhaustive.

^{*} Corresponding authors. E-mails: Khodair_62@yahoo.com, Najim.Al-Masoudi@gmx.de

[#] Present address: Riaydh Teachers College, P.O. Box 4341, Riaydh 11491, Saudi Arabia

[§] Present address: EuroMed-Konstanz, P.O. Box 100552, D-78405 Konstanz, Germany



Several thiohydantoins exhibited a variety of biological activities including antiviral and antitumor activities. Examples of such compounds were 5-(2-thienylmethylene)-3-phenyl-2-(2,3,4,6-tetra-*O*-acetyl- β -*D*-glucopyranosyl)-2-thiohydantoin (**5a**) and the 3-(4-chlorophenyl) analogue **5b** [10], which showed remarkable activity against both HSV-1 and HSV-2. The thioglycosyl hydantoin **6a** proved to possess a broad spectrum antitumor activity against a wide range of different human cell lines of nine tumor subpanels causing both cytostatic and cytotoxic effects, with medium lethal concentrations (*LC*₅₀) at 15.1, 41.7, and 83.2 μ M. On the other hand, the galactoside analogue **6b** showed potential selectivity against leukemia cell lines [11] (Scheme 2).

Attention had been focused on the synthesis of glycosylated hydantoin derivatives because of their resemblance with natural nucleosides, in particular (E)-l-(2deoxy- β -D-erythropentofuranosyl)-5-ethylidene-2,4-imidazolidinedione (7a) [12], a potent nucleoside derived from thymidine (8). It is derived by ring contraction of the nucleobase by a *de facto* rearrangement of carbonyl from C-5 to C-6. Thymidine itself was isolated a long time ago from thymonucleic acid [13-16]and consequently there has been strong interest in synthesizing modified thymine nucleosides as potential drugs. The possibility of finding interesting five-membered ring analogues of thymidine is best illustrated with showdomycin (9), which is a broad spectrum antibiotic first isolated from *Streptomyces showdoensis* [17]. This antibiotic has been found to exhibit activity against *Ehrlich ascites* tumor in vivo and against cultured HeLa cells [18, 19]. Suhadolink et al. [20] and Kozikowski et al. [21] have reported the antibacterial and antitumor activities of the similar class of the C-nucleosides family [20, 21] (Scheme 3). Recently, Dewynter et al. [22] described the preparation of a series of pseudo-nucleosides containing the sulfahydantoin 1,2,5-thiadiazolidine-3-one-1,1-dioxide (10) [23] as aglycone.

Azidothymidine (*AZT*, Zudividine[®], **11**) [24] was the first drug at the market to treat HIV infection. However, it displays remarkable toxicity [25, 26]. Therefore,



there is still an urgent need for new antiviral agents with low toxicity to normal cells. Such compounds might be *AZT* analogues bearing 2,4-imidazolidindione as a nucleobase, *e.g.* (*E*)- and (*Z*)-1-(3-azido-2,3-dideoxy- β -*D*-*erthyro*pentofuranosyl)-5-(substituted ethylidene)-2,4-imidazolidinediones **12** and **13** [27].

2-Thiohydantoin Nucleosides

Nitrogen Glycosylation Reactions

N-3 Glycosylation involves the reaction of glycine ethyl ester hydrochloride nucleoside with pyranosyl sugars. The main synthetic route to 2-thiohydantoin



nucleosides started with the construction of the heterocycle residue from suitably C-1 functionalised sugar derivatives. Reaction of 2,3,4,6-tetra-*O*-acetyl- β -*D*-glucopyranosyl isothiocyanate (**15**) [28, 29], obtained form the bromo analogue **14** and silver thiocyanate with glycine ethyl ester hydrochloride in the presence of pyridine, gave almost quantitatively ethyl (2,3,4,6-tetra-*O*-acetyl- β -*D*-glucopyranosyl) thiohydantoate (**16**). The oxygen analogue **17** was prepared from **16** by treatment with alcoholic silver nitrate. Deblocking of **16** with *Me*OH/HCl followed by saponification and acidification with hot HCl furnished **18**. Similarly, **19** was obtained from **17**. Meanwhile **20** and **21** were prepared from saponification of **16** and **17** followed by acidification with cold HCl.

Fernandez et al. [30] have applied the same approach for the cyclization of the thiourea residue in the presence of base to furnish the 2-thiohydantoins. Thus, treatment of *N*-ethoxycarbonylmethyl-*N'*-(2,3,4,6-tetra-*O*-acetyl- β -*D*-glucopyranosyl)thiourea (16) or the - β -*D*-ribopyranosyl analogue 22 with KOH, followed by acidification furnished 3-(β -*D*-glucopyranosyl)-2-thiohydantoin (18) or the ribo analogue 23 (Scheme 6).

Reaction of the glucopyranosylamine hydrobromide **24** and hydrochloride analogue **25**, prepared from the corresponding 2,3,4-tri-*O*-acetyl-1-*N*-(2,2-diethoxy-carbonylvinyl)-6-*O*-trityl- β -*D*-gluco-pyranosylamines [31] hydrobromide (hydrochloride) with thiophosgene in basic medium afforded the isothiocyanate derivatives **26** and **27** [32]. Treatment of **26** and **27** with amino acetone hydrochloride followed by deacylation gave the 2-thioimidazoline derivative **28** [33] (Scheme 7).



Scheme 6



Sulfur Glycosylation Reactions

Thioglycosides of 2-Thiohydantoins Carrying Pyranosyl Sugars

The main synthesis route to thiohydantoin carrying glycosides was the direct condensation between the appropriate 2-thiohydantoin bases with the bromo peracylated sugars in the presence of aqueous base. Treatment of 5-arylidene-3-phenyl-2-thiohydantoins 29a-29e with 14 in the presence of aqueous NaOH and acetone at 23°C provided the thioglycosides 30a-30e, while hydrolysis of 30a with conc. *Me*OH/HCl afforded the free bases 29a and 31 [34] (Scheme 8).

Following the same approach, 3-substituted-2-thiohydantoins **32a**–**32d** were reacted with **14**, the *D*-galactopyranosyl- **33**, and *D*-xylopyranosyl-analogues **34** to furnish, probably *via* Walden inversion [35, 36], the corresponding 5-arylidene-3-substituted-2- β -*D*-glucopyranoside (**35a**–**35d**), β -*D*-galactopyranoside (**35e**–**35h**), and β -*D*-xylopyranoside (**35i**–**35m**) derivatives of 2-thiohydantoins [37] (Scheme 9).

Numerous examples of thioglycosides carrying 2-thiohydantoins with various substituents have been synthesized in the presence of hydride ions. Thus, 5a-5g were obtained from treatment of 36a-36g with 14 in the presence of NaH. Alternatively, 5g has been synthesized by treatment of 3-phenyl-2-thiohydantoin 37 with aqueous NaOH and acetone [10] (Scheme 10).

When 5-arylidene-3-substituted-2-thiohydantoins 38a-38d were treated with 1.1 eq. of aqueous NaOH or K₂CO₃ in acetone followed by addition of 14, the corresponding (*Z*)-2-thiohydantoin nucleosides 39a-39d were obtained. Hydrolysis of 39a or 39c with conc. HCl in refluxing *Et*OH afforded (*Z*)-5-arylidene-3-



substituted-2,4-imdazolidindiones **40a** or **40b**. Following the same method, the galacto analogues **39e–39h** were synthesized by condensation of **38a–38d** with **33** [38] (Scheme 11).

2-Thiohydantoin derivatives **41a**–**41d** bearing a heterocyclic side chain at position 5 were reacted with **14** or the galacto analogue **33** in the presence of hydride ions and *Me*CN as the solvent to give 5-arylidene-3-substituted-2-thiohydantoin nucleosides **6a**, **6c**, and **6d**, as well as the galacto derivative of 2-thiohydantoin **6b** [39] (Scheme 12). More examples of the gluco and galacto derivatives of 2-thiohydantoins having differently substituted 5-(Z)-arylidene groups, such as 3-phenyl-, 3-methyl-, 2-thienyl-, 2- and 3-indolyl-, 2-pyridyl-, and furyl-groups, have been reported [40, 41]. These new derivatives of the thioglycosides **43** bearing 3-allyl-5-(Z)-arylidene groups were prepared by condensation of the corresponding 2-thiohydantoins **42a**–**42f** with **14** in presence of hydride ions in anhyd. *Me*CN. Compound **43a** was synthesized *via* an alternative route by condensation of the silyl



derivative 44, which in turn was prepared from 42a with bis-(trimethylsilyl)acetamide (BSA) in MeCN, and α -D-glucose pentaacetate (45) in presence of trimethylsilyl trifluoromethanesulfonate (TMSTf). The thioglycoside 43a was isolated by



Scheme 13



chromatography and no *N*-nucleoside was detected [41] (Scheme 13). In a similar manner, the galacto-analogues have been prepared [42].

Thioglycosides of 2-Thiohydantoins Carrying Furanosyl Sugars

Pedersen et al. [10] have followed the previous synthetic routes to prepare the thioglycosides carrying furanosyl residues, such as 5-benzylidene-3-phenyl-2- $(3,5-di-O-toluoyl-erythro-\beta-D$ -pentofuranosyl)-2-thiohydantoin (**47a**) and its 5-(2-thienyl)-analogue **47b**, by condensation of the corresponding bases **29a** and **36a** with **46** in the presence of NaH in anhyd. *Me*CN (Scheme 14). Similar treatment of **42a**, **42b** with **48** afforded the thioglycosides **49a**, **49b**. Alternatively, **49a** was synthesized in a good yield by condensation of the silylated base **44** with **50** in the presence of *TMSTf* and anhyd. *Me*CN [41] (Scheme 14).

Nitrogen and Sulfur Glycosylation Reactions

Bis-glycosylation takes place on both *S*- and *N*-atoms of the unsubstituted hydantoins. The bis-glycosylated hydantoin was hydrolysed to the *N*-3 glycosylated analogue on treatment with NH₃/*Me*OH. Thus, reaction of 5-arylidene-2-thiohydantoins **51a** or **51b** with **14** in the presence of aqueous K₂CO₃ afforded **52a** or **52b**, which gave the 5-(*Z*)-arylidene-3-(β -*D*-glycopyranosyl)-hydantoins **53a** or **53b** on treatment with NH₃/*Me*OH [40] (Scheme 15). Similarly, the *D*-galacto analogues have been prepared [38].

Various derivatives of bis-glycosilated derivatives of 2-thiohydantoins 2 have been synthesized by reaction of 2 with aromatic aldehydes in presence of ethanolic K_2CO_3 followed by addition of 14 in acetone to give 54a, 54b. Deprotection with



Scheme 16

NH₃/*Me*OH afforded 5-(*Z*)-arylidene-3-(β -*D*-glycopyranosyl)hydantoins 55a, 55b [40] (Scheme 16).

When 5-arylidene-2-thiohydantoins **56a–56c** were treated with **14** in aqueous acetone, mono- and bis-glycosylation afforded **57a–57c** and **58a** or **58b**. Upon deprotection of the bis-products **58a** or **58b** with NH₃/*Me*OH, the thioglycosyl group was most likely replaced by an oxo group *via* a nucleophilic substitution reaction. Subsequent deacetylation afforded the *N*-3 glycosyl hydantoin derivatives **59a** or **59b** [10]. However, formation of the 2-oxo derivative directly from the moisture cannot be excluded. This type of cleavage explains why the deprotection of compounds **5** [10], **30a–30e** [34], **35a–35m** [37], **39a–39h** [38], **6a–6d** [39], **43a–43l** [41], **47a** or **47b** [10], and **49a** or **49b** [41] in saturated NH₃/*Me*OH is unsuccessful. Recently, various derivatives of such nucleosides have been prepared [42–44].

The glycosylation of the hydantoins was confirmed by comparison with the previously reported ¹H NMR (*DMSO*-d₆) spectrum of 5-(phenylmethylene)-3methylhydantoin, which has been characterized by the signal of N¹-H at $\delta = 10.72$ ppm whereas N³-H of the 1-methylhydanoin analogue resonated at $\delta = 11.38$ ppm [45]. The ¹H NMR spectra of **59a** and **59b** showed an NH signal at $\delta = 10.9$ and 10.70 ppm corresponding to N¹-H, proving the N-3 glycosylation. The glycosylation at N-3 of **59a** was further confirmed by NOE experiments. Thus, on irradiation at 10.91 ppm, a large NOE enhancement was found for the *ortho* protons a small enhancement was detected for NH. These data proved the (*Z*)-configuration of the exocyclic double bond in **59a** [10] (Scheme 17).



Scheme 17

Glycosylation Reactions Involving 5-(*Uracil-5-ylmethylene*)-2-*Thiohydantoin with Furanosyl Sugars*

5-Formyluracil (60) prepared in two steps from uracil in 30% yield according to Refs. [46, 47] was condensed with 2 or 3-phenyl-2-thiohydantoin (37) by refluxing in a mixture of NaOAc and HOAc to give 5-(uracil-5-ylmethylene)-2-thiohydantoin (61a) and the 3-phenyl-analogue 61b. Silylation of 61a or 61b with hexamethyl-disilazane (*HMDS*) in presence of (NH₄)₂SO₄ as the catalyst [48] followed by addition of 62 [49, 50] in the presence of *TMSTf* [51] as a *Lewis* acid and anhyd. *Me*CN as the solvent afforded an anomeric mixture (α/β) of the corresponding 5-[1-(2-deoxy-3,5-bis-O-(4-methylbenzoyl)-*D-erythro*-pentofuranosyl)uracil-5-ylmethylene]-2-thiohydantoins 64a, 64b. The glycosylation took place on the pyrimidine and not on the hydantoin ring. This was proven by synthesizing the same nucleosides by an alternative route. Thus, coupling of the silylated derivative of 60 with 62 afforded a mixture of 63 in 63% (α : β = 1:2). Condensation of 63 with 2 and 37 by using piperidine as the catalyst provided the α - and β -anomers of 64a and 64b. Deprotection with NaOMe in MeOH afforded the free nucleosides 65a and 65b, which were separated by chromatography [52] (Scheme 18).

Similar treatment of the silvlated 5-formyluracil **60** with **66** resulted in the formation of the uracil nucleoside **67** (72%). Coupling of **67** with **2** or **37** by using piperidine as the catalyst gave the corresponding α/β mixture of 5-(2,3,5-tri-*O*-acetyl-*D*-ribofuranosyluracil-5-ylmethylene)-2-thiohydantoin (**68a**) and the 3-phe-nyl analogue **68b**. Only one diastereoisomer of **68** with regard to the new double bond was observed, which most likely is of (*Z*)-configuration as compared to the products obtained from condensation of hydantoins with aldehydes [42]. However, confirmation was not possible because the (*E*)-stereoisomer was not available as a reference. Removal of the acetyl groups of **68a** and **68b** by NaOMe in MeOH



Scheme 18



Scheme 19



furnished after chromatography the 5-(β -*D*-ribofuranosyluracil-5-ylmethylene)-2thiohydantoins **69a** and **69b**. An alternative route for the synthesis of **68a** involved condensation of the silylated **61a** with the sugar moiety **66** in presence of *TMSTf*. The β -anomer was isolated by chromatography [53]. The NOE investigation of **68b** showed a strong enhancement for H-6 on the uracil ring on irradiation of H-2'. This indicated a preferential *anti*-rotamer conformation of the nucleobase around the glycosidic bond [53] (Scheme 19).

2,4-Dithiohydantoin Nucleosides

5,5-Dimethyl-3-morpholinomethyl-2-(2,3,4,6-tetra-*O*-acetyl- β -*D*-hexopyranosyl)-2,4-dithiohydantoins (**71a** and **71b**) and the xylopyranosyl analogue **71c** were synthesized by the reaction of 5,5-dimethyl-3-morpholinomethyl-2,4-dithiohydantoin (**70**) with **14**, **33**, and **34** in presence of aqueous KOH and acetone at 23°C [37] (Scheme 20).

Hydantoin Nucleosides

Nitrogen Glycosylation Reactions Involving Hydantoins with Furanosyl Sugars

N-1 Glycosylation Reactions

It has been reported that silvlation of hydantoin bases occured on both amido groups to give the bis- silvlated derivatives, such as the silvlated 5-alkylidene-**73a** and 5-arylidene-2,4-imidazolidinediones **73b**–**73f** (Scheme 21). Various hydantoin nucleosides have been synthesized by coupling the silvlated hydantoins with the sugar precursors in the presence of a *Lewis* acid. For example, 1-(2-deoxy-3,5-di-*O*-(4-methylbenzoyl)-*D*-*erythro*-pentofuranosyl)-5-(substituted methylene)-2,4-imiazolidinediones **74a**–**74d** were obtained in 60–80% (*E*/*Z*), as reported by *Vorbrüggen et al.* [51, 54], by coupling of **73a**–**73d** with **62** in presence of *TMSTf*. No other nucleoside was detected in this reaction. Removal of the toluoyl group of **74a**–**74d** with NaOMe in MeOH at 23°C furnished the free nucleosides (*E*/*Z*)





7a–7d and **75a–75d** [12] (Scheme 22). Unexpectedly, treatment of **74c** with NH₃/*Me*OH at 23°C showed incomplete reaction even after 3 days, and instead the three partially protected nucleosides **76–78** were isolated. Characteristic downfield shifts were observed for H-5' in the ¹H NMR spectra of the α/β mixture **77** and **78** in comparison with those of H-5' of the 5'-*O* deprotected **76** [12] (Scheme 23).

When the trimethylsilyl derivative **73e** was condensed with **62** in the presence of *TMSTf* at -10° C for 3 days, two products were isolated, namely (*Z*)-l-(2deoxy-3,5-bis-*O*-(4-methylbenzoyl)- β -*D*-*erythro*-pentofuranosyl)-5-(1-naphthylmethylene)-2,4-imidazolidinedione (**79**) and 7-[2,4-bis-*O*-(4-methylbenzoyl)-ldeoxy-*D*-erythrit-1-yl]benz[*f*]imidazo[1,5-*b*]isoquinoline-9,11-(7*H*,10*H*)-dione (**80**). These were separated by chromatography in 20 and 50% yields. The free





nucleosides **81** and **82** were obtained by treatment with NaOMe in MeOH at 23° C [12] (Scheme 24).

Additional hydantoins bearing different substituents were prepared such as the (2S,3S)-1,3-bis-O-(4-methylbenzoyl)-5,5-bis[5-[2,4-dioxo-5-imidazolidinylidene)methyl]furan-2-yl]pentane-1,2,3-triol (83) (21%) from coupling of the trimethylsilyl derivative 73f with 62 in presence of *TMSTf* at -10° C for 3 days. Removal of the protecting toluoyl groups from 83 with NaOMe in MeOH at 23°C furnished 84 [12] (Scheme 25).

Interesting examples of hydantoin nucleosides were synthesized by coupling the azido sugar derivative **85** [55] with the silylated bases **73a**, **73b** in presence of *TMSTf* to give the 1-(3-azido-5-*O-tert*-butyldiphenylsilyl-2,3-dideoxy-*D-erythro*pentofuranosyl)-5-(alkylmethylene)-2,4-imidazolidinediones **86a**, **86b**. The nucleoside **86a** was isolated as a 4:1 (α : β) anomeric mixture (E/Z) in 46% yield, while **86b** was isolated as a 4:1 (α : β) mixture (50%). Deprotection of **86a** with tetrabu-



Scheme 25

tylammonium fluoride in *THF* at 23°C provided the (*E*)-l-(3-azido-2,3-dideoxy- β -*D*erythro-pentofuranosyl)-5-ethylidene-2,4-imidazolidinedione (**12a**) (17%) and the (*Z*)-isomer **13a** (25%). In a similar manner, (*Z*)-l-(3-azido-2,3-dideoxy- β -*D*erythro-pentofuranosyl)-5-(phenylmethylene)-2,4-imidazolidindione (**13b**) and the α -anomer **13c** were obtained from **86b** in 2% and 38% yields [27] (Scheme 26).



Scheme 26



Scheme 27

Pedersen et al. [56] have described the synthesis of (*Z*)-2,4-imidazolidinedione nucleosides **88** (22%) as $\alpha:\beta$ (1:1) mixture by coupling of **87** with **73b** at -30° C in presence of *TMSTf*. Removal of the toluoyl group of **88** was employed by treatment with NaOMe in MeOH at 23°C to give (*Z*)-1-(5-azido-2,5-dideoxy-*D*-erythro-pentofuranosyl)-5-(phenylmethylene)-2,4-imidazolidinediones (**89**), which were separated by chromatography. Reduction of **89** β with tin(II) chloride in MeOH afforded the corresponding amino analogue **90** β in 10% yield (Scheme 27).

Similarly, condensation of **73b** with the sugar derivative **91** [57] in presence of *TMSTf* at -30° C yielded the $\alpha:\beta$ (4:5) mixture of (*Z*)-1-(5-*O*-acetyl-2,3-dideoxy-3-phthalimido-*D*-erythro-pentofuranosyl)-2,4-imidazolidinediones (**92**) (22%), which were isolated by chromatography. Removal of the phthaloyl group was performed of both α - and β -anomers with methylamine in abs. *Et*OH at 23°C to furnish the free nucleoside **93** β as well as **93** α (Scheme 28).

N-3 Glycosylation Reactions

Muramatsu et al. [58, 59] have reported the formation of 1-glycosylated 5-hydroxyhydantoins **94a** and **94b** by ozonolysis of cytidine, uridine, and thymidine as



Scheme 29

examples of *N*-3-glycosyl hydantoin nucleosides. These were followed by syntheses of interesting nucleosides, such as 3-(2,3,5-tri-*O*-benzoyl- β -*D*-ribofuranosyl)-5-(carbomethylidene)hydantoin (**97**), which was obtained predominately by coupling of the silylated hydantoin **95** with **96** using SnCl₄ as *Lewis* acid and CH₂Cl₂ as the solvent. Debenzoylation of **97** with NaOMe in MeOH yielded the corresponding free nucleoside **98** [60] (Scheme 29).

Nitrogen Glycosylation Reactions Involving Hydantoins with Pyranosyl Sugars

Mentzer et al. [61] have synthesized 5-alkylidene-3-(β -D-glucopyranosyl)hydantoins **99a** and **99b** by coupling 5-alkylidenehydantoins **72a** and **72g** with **14** in the presence of HgCl₂ and NaOH (Scheme 30).

Powell et al. [62, 63] have reported the synthesis of β -*D*-glucopyranosylnitrohydantoin **101** by dissolving **100** and benzylethylammonium bromide in aqueous NaOH followed by treatment with a solution of **14** in CHCl₃. Compound **101** was also prepared by an alternative method from **100** and **14** under phase transfer catalyst conditions [64]. More examples in the series of *N*-3 hydantoin nucleosides bearing pyranosyl sugars have been reported, such as the 3-(β -*D*-glucopyranosyl)-5,5-diphenylhydantoin (**102**) [64] (Scheme 31).



Scheme 31

References

- [1] Baeyer A (1861) Liebigs Ann Chem 117: 178
- [2] Klason P (1890) Chem Ztg 14: 543; cf also Tiwari SS, Swaroop A (1963) J Ind Chem Soc 40: 693
- [3] Vigneaud VD, Melville DB (1949) The Chemistry of Penicillin. In: Clarke HT, Johnson JR, Robinson R (eds) Princeton Univ, p 269
- [4] Johnson TB, Chemoff LH (1912) J Am Chem Soc 34: 1208
- [5] Cook AH, Heilbron I, Levy AL (1948) J Chem Soc 201
- [6] Ware E (1950) Chem Rev 46: 403
- [7] Schipper ES, Day AR (1957) Heterocyclic Compounds. In: Elderfield RC (ed), vol. 5. Wiley, New York, p 254
- [8] Edward JT (1966) Chem Org Sulfur Comp 2: 287
- [9] Lopez CA, Trigo GG (1985) Adv in Heterocycl Chem 38: 177
- [10] El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C (1994) J Med Chem 37: 73
- [11] Al-Obaid AM, EL-Subagh HI, Khodair AI, Elmazar MMA (1996) Anticancer Drugs 7: 873
- [12] El-Barbary AA, Khodair AI, Pedersen EB (1993) J Org Chem 58: 5994
- [13] London L (1929) J Biol Chem 83: 793
- [14] Tipson L (1935) J Biol Chem 109: 623
- [15] Lemieux RU (1961) Can J Chem 39: 116
- [16] Tollin P, Wilson HR, Young DW (1968) Nature 217: 1148
- [17] Nishumura H, Mayama M, Komatsu Y, Kato H, Shimaoka N, Tanaka Y (1964) J Antibiot 77: 148
- [18] Matsuura S, Shiratori O, Katagiri K (1964) J Antibiot 17: 234
- [19] Nakagawa Y, Kano H, Tsukuada Y, Koyama H (1967) Tetrahedron Lett 4105
- [20] Suhadolnik RJ (1970) Nucleoside Antibiotics. Wiley-Interscience, New York, p 354
- [21] Kozikowski AP, Ames A (1981) J Am Chem Soc 103: 3923
- [22] Dewynter G, Aouf N, Regaini Z, Montero J-L (1996) Tetrahedron 52: 993
- [23] Aouf N, Dewynter G, Montero J-L (1991) Tetrahedron Lett 32: 6545
- [24] Mitsuya H, Weinhold KJ, Furman PA, St. Clair MH, Lehrman SN, Bolognesi RC, Barry DW, Broder S (1985) Proc Natl Acad Sci USA 82: 7096
- [25] Nasr M, Cradock J, Johnston MI (1992) AIDS Res Human Retrovir 8: 135
- [26] De Clercq E (1992) AIDS Res Human Retrovir 8: 119
- [27] El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C (1994) Arch Pharm 327: 653
- [28] Fisher E (1914) Ber Deutsch Chem Ges 47: 1382
- [29] Haring KM, Johnson TB (1933) J Am Chem Soc 55: 39530
- [30] Mota JF, Fernandez JMG, Adeian MAP, Mellet CO, Gomez MG (1990) Ann Quim 86: 655; Chem Abstr (1991) 114: 122900m
- [31] Fuentes J, Pradera MA, Robina I (1991) Tetrahedron 47: 5797, and references therein
- [32] Babiano R, Fuentes J, Galbis JA (1986) Carbohydr Res 154: 280
- [33] Fuentes J, Moreda W, Ortiz C, Robina I, Weish C (1992) Tetrahedron 48: 6413
- [34] El-Barbary AA, Saffan AA, Khodair AI (1990) Delta J Sci 14: 601; Chem Abstr (1992) 117: 171851s
- [35] Wood HB, Fletcher HG (1956) J Am Chem Soc 78: 207
- [36] Bonner WA (1959) J Am Chem Soc 81: 1448
- [37] Aly YL, El-Barbary AA, Hashem AFM, El-Shehawy AA (2004) Phosph Sulfur Silicon 179: 185
- [38] Khodair AI, Ibrahim EI (1996) Nucleos Nucleot 15: 1927
- [39] Khodair AI, El-Subbagh HI, El-Emam AA (1997) Boll Chim Farm 136: 561
- [40] Khodair AI (1997) Phosph Sulfur Silicon 122: 9
- [41] Khodair AI, Gesson J-P (1998) Phosph Sulfur Silicon 142: 433
- [42] Khodair AI (2001) Carbohydr Res 331: 445
- [43] Khodair AI, El-Barbary AA, Abbas YA, Imam DR (2001) Phosph Sulfur Silicon 170: 261
- [44] Khodair AI (2001) Nucleos Nucleot & Nucleic Acids 205: 1735

- [45] Tan SF, Ang KP, Fong YF (1986) J Chem Soc Perkin Trans II 194
- [46] Cline RE, Fink RM, Fink K (1959) J Am Chem Soc 81: 2521
- [47] Ressner EC, Fraher P, Edelman MS, Martes MP (1976) J Med Chem 19: 194
- [48] Wittenburg E (1964) Z Chem 4: 303
- [49] Motawia MS, Pedersen EB (1990) Liebigs Ann Chem 599
- [50] Hoffer M (1960) Ber Deutsch Chem Ges 93: 2777
- [51] Vorbrüggen H, Krolikiewics K, Bennua B (1981) Ber Deutsch Chem Ges 114: 1234
- [52] El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C (1994) Monatsh Chem 125: 593
- [53] El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C (1994) Liebigs Ann Chem 619
- [54] Vorbrüggen H, Hofle G (1981) Ber Deutsch Chem Ges 114: 1256
- [55] Hansen P, Pedersen EB (1990) Acta Chem Scand 44: 522
- [56] El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C (1994) Nucleos Nucleot 13: 707
- [57] Motawia MS, Wengel J, Abdel-Megied AES, Pedersen EB (1989) Synthesis 384
- [58] Matsui M, Nakazumi H, Kamiya K, Yatome C, Shibata K, Muramatsu H (1989) Chem Lett 723
- [59] Matsui M, Inoue T, Shibata K, Muramatsu H (1990) Bull Chem Soc Jpn 63: 296
- [60] Mikailopulo IA, Kalinichenko EN, Akhrem AA (1981) J Carbohydr Nucleos Nucleot 8: 277
- [61] Guyot PA, Chopin J, Mentzer C (1960) Bull Soc Chim France 1596
- [62] Ahmed B, Powell JW (1990) J Chem Soc Pakist 12: 105; Chem Abstr (1991) 114: 6986v
- [63] Ahmed B, Powell JW (1989) Ind J Pharm Sci 51: 59; Chem Abstr (1990) 112: 99024b
- [64] Cheh CH, Ta CM (1989) Chung-Hua Yao hsueh Tsa Chih 41: 189; Chem Abstr (1990) 112: 119257k